



Full Length Research Paper

# Functional properties change of Nagara bean flour through fermentation by *Lactobacillus plantarum*

\*Susi Susi<sup>1</sup>, Lya Agustina<sup>1</sup> dan Condro Wibowo<sup>2</sup>

<sup>1</sup>Department of Agroindustrial Technology, Lambung Mangkurat University Banjarbaru South Kalimantan, Indonesia

<sup>2</sup>Department of Food Science and Technology, Jenderal Soedirman University, Purwokerto, Central Java Indonesia

\*Corresponding Author Email: [susi\\_tip@unlam.ac.id](mailto:susi_tip@unlam.ac.id)

## ABSTRACT

Nagara bean (*Vigna unguiculata* ssp. *Cylindrica*) is one of cowpea subspecies which has a high carbohydrate content of 50-60% and 20-25% protein. In order to increase flour products of Nagara cowpea, its functional properties can be improved through fermentation by using *Lactobacillus plantarum*. This research was examined functional properties changes of Nagara bean flour fermented by *L. plantarum* from 24 to 120 hours using their size variation of a whole and grits bean. The best result of degree of water absorption, swelling volume and gelatinization of cowpea flour were showed along 48 hours fermentation and grits size with 198.29% level of water absorption db, peak viscosity of 2500 cP and final viscosity of 2500 cP respectively.

**Keywords** : Nagara bean, water absorption, swelling volume, solubility, gelatinization profile

## INTRODUCTION

Nagara bean (*Vigna unguiculata* ssp. *Cylindrica*) is a local cowpea grown in South Kalimantan and well adapted both in dry lands and lowland swamps. Olapade *et al.* (2012) stated that cowpea contained a high carbohydrate of 50-60% and 20-25% of protein with dominant amino acids classified as arginine, isoleusine, glutamate and aspartate (Tangadurai, 2005). Susi (2012) researched that dominant essential amino acids of cowpea were 0.734% valine, 0.791% metionine and 0.775% phenilalanine; and dominant non essential amino acids were 0.913% aspartic acid, 2.182% glutamic acid and 0.826% histidine. Moreover, nutrient of cowpea was suitable for people who were not consuming meat and fish (Atuobi *et al.* 2011).

The useful of Nagara bean in Kalimantan is limited only for additional food in cooking. Increase of Nagara bean use can be achieved by knowing functional properties of Nagara bean flour. Niba (2003) reported limitation of legume characteristics could be improved by applying bioprocess and fermentation technique. This bean has anti nutrients consisted of 1.9g/100 g total phenolic compound and 5.6g/100g tannin (Thangadurai 2005). Fermentation technique is usually used for improving quality of starch such as structure modification,

physical and chemical characteristics (Chinsamran *et al.* 2005), whereas this technique influences solubility properties, swelling volume and flour viscosity of products used for next processing.

The research aimed to examine functional properties of Nagara bean flour through fermentation processes by *Lactobacillus plantarum* using variation in periods fermentation and in grain size. Lactic acid bacteria like *L. plantarum* functions as fermentation agents of flour to modify the functional characteristics. These bacteria classified as Amilolitic-Lactic Acid Bacteria have a capability to degrade flour both in direct ways and in facultative hetero-fermentative methods. The functional characteristics of Nagara bean were investigated degree of water absorption, swelling volumes and gelatinization profiles.

## MATERIALS AND METHODS

### Materials

Nagara bean was originated in Nagara – Hulu Sungai Selatan district of South Kalimantan. A water bath

shaker, a centrifuge, an oven and a glassware were used in the research.

### Spontaneous Fermentation Process

A total of fermentation using Lactic Acid Bacteria *L. plantarum* was 1% (v/v) with the size of bean were a whole grain and grits. The ratio of Nagara bean and water soaking was one fourth with different fermentation periods for instance 24, 48, 72, 96 and 120 hours. Soon after fermentation, Nagara bean was rinsed and peeled and then dried in 60°C for 48 hours. Next, the bean was floured for 80 mesh.

Data analysis was done for water absorption, swelling volume, solubility and gelatinization profile by using Rapid Visco Analyzer Model RVA-S4 with the Thermocline for Windows (TWC) program. In addition, granular characteristics were examined by Scanning Electron Microscopy (SEM).

### Water Absorption Capacity by gravimeters

A centrifugal tube was filled 2 g flour (a) and added distilled water and spun with a vortex. After that, the sample was waited for 30 minutes and centrifuged with the speed 3000 rpm for 15 minutes, and stored in the desiccators then weighed (b).

Water Absorption Capacity (%db)

$$\text{Water absorption capacity (\%db)} = \frac{b - a}{ms} \times \frac{100\%}{(1 - ka)}$$

, whereas

a = dried sample + centrifugal tube (g)

b = wet sample + centrifugal tube (g)

ms = sample (g)

### Solubility dan Swelling volume

Swelling volume was examined by weighting 0.35 g of flour and added 12.5 mL water and placed in a centrifugal tube. After that, the sample was spun by a vortex and placed in waterbath for 92.5°C and then every 5 minutes was vortexed for 10 minutes. The sample was cooled in an ice for 1 minutes and warmed 25°C for 15 minutes. Then, the sample was centrifuged for 3600 rpm for 15 minutes. The volume of formed gel was measured (swelling volume) in mL/g (db). The solubility was formed by pouring supernatant to weighted cup and then dried for 110°C overnight.

$$\text{Solubility} = \frac{wl}{wdm} \times 100\%$$

$$\text{Swelling volume (ml/g bk)} = \frac{w2}{wdm} \times 100\% ;$$

whereas

wdm = ws (1-mc)

w1 = supernatant (g)

w2 = volume of formed gel (mL)

ws = sample (g)

mc = water level (desimal) of flour in wet basis

## RESULTS AND DISCUSSION

### Water Absorption Capacity

Granular starch is the main component of flour which can swell in cool or hot water. In cool water, the capacity of water absorption is limited; however, when the starch is heated the formed energy can break hydrogen bonds causing the area of water absorption enlarges and increases its capacity.

Water absorption capacity of control (non fermented Nagara bean flour) was 151.45% and fermentative treatments could increase the capacity of water absorption. Analysis of variance ( $\alpha = 5\%$ ) showed the significant differences in fermentation periods to the capacity of water absorption of fermented Nagara bean flour. Table 1 explained water absorption of Nagara bean flour.

Water absorption of fermented Nagara bean indicated that an increase of water absorption was in line with fermentation periods until 120 hours for whole beans; nevertheless it was not affected by fermentation periods for 96 and 72 hours.

Another study reported that the activity of proteolytic microorganism influenced added polar groups of granular starch along fermentation processes and these groups could lift hydrophilic flour (Etudaiye *et al.* 2009).

### Swelling Volume

A swelling volume reveals degree of swelling granular starch when interacts with water. The more water is absorbed, the increase of swelling granular starch happens. The percentage of swelling volume of non fermented Nagara bean flour was 590.48% and adding *L. plantarum* as a fermentative agent produced an increase of swelling volume. Analysis of variance ( $\alpha=5\%$ ) noted that the single fermentation periods influenced swelling volume of flour. Table 1 illustrates the swelling volume.

A swelling volume of fermented Nagara bean flour by *L. plantarum* increased in fermentation periods for 72 hours both a whole bean and grits; however, it decreased after 72 hours of fermentation. Swelling volume of flour is usually related to protein and starch of flour. Along fermentation processes, Lactic Acid Bacteria (LAB) produce proteolytic enzymes functioning to degrade protein in Nagara bean. Moreover, LAB also produce amylase to degrade amylose, so amylose content

**Table 1.** Water absorption, swelling volume and solubility of fermented Nagara bean flour by *L. plantarum*

Treatments		Parameters			
Size	Fermentation (hours)	periods	Water absorption (% db)	Swelling volume (% db)	Solubility (% db)
Whole	24		160.87±23.56 <sup>a</sup>	622.05±38.88 <sup>b</sup>	24.67±0.17 <sup>a</sup>
	48		192.19±1.45 <sup>b</sup>	714.61±3.30 <sup>c</sup>	7.76±1.47 <sup>b</sup>
	72		211.58±8.01 <sup>bc</sup>	662.79±73.74 <sup>b</sup>	5.54±0.94 <sup>b</sup>
	96		211.38±1.08 <sup>c</sup>	614.93±44.46 <sup>ab</sup>	6.16±0.21 <sup>b</sup>
	120		215.07±3.89 <sup>c</sup>	567.33±99.72 <sup>a</sup>	5.96±0.92 <sup>b</sup>
Grits	24		197.16±5.53	712.35±55.43	20.52±2.80
	48		198.29±2.98	739.79±5.18	6.00±0.09
	72		202.88±2.33	646.06±33.35	5.96±0.01
	96		220.41±1.19	623.34±17.08	5.92±0.12
	120		216.92±11.14	586.28±87.72	5.91±0.14

\*: the same letter indicates duncan test which is not significantly different

**Table 2.** The gelatinization profiles of fermented Nagara bean flour

Treatments		Gelatinization profile of Nagara bean flour							
Size	Fermentation time (hours)	Peak visc (cP)	Trough visc (cP)	Breakdown visc (cP)	Final Visc (cP)	Seat back visc (cP)	Peak Time (min)	Peak Temp (°C)	Pasting temp (°C)
Whole	24	1845	1137	708	1668	531	7.80	91.00	79.30
	48	2453	1159	1294	2124	965	7.73	90.65	78.05
	72	2633	1306	1327	2460	1154	7.93	91.85	77.70
	96	2301	1163	1138	2284	1121	8.00	92.20	78.05
	120	2349	1249	1100	2427	1178	8.33	94.20	78.85
Grits	24	2322	1140	1182	2123	983	7.80	91.00	78.05
	48	2722	1326	1396	2500	1174	7.87	91.45	78.10
	72	2713	1322	1391	2650	1328	7.93	91.85	77.70
	96	2438	1181	1257	2499	1318	8.13	92.95	78.50
	120	2563	1299	1264	2906	1607	8.13	93.00	78.10
Control		2093	1074	1019	1617	543	7.53	89.30	78.10

decreased. The low amylosa can enhance swelling volume of starch (Tester and Morisson, 1990). Leach *et al.* (1959) explained that swelling volume was influenced by granular bonding. LAB yielded amylase attacking amorphous area of starch, and then LAB hydrolytic performances were induced by both enzymes and organic acids produced in fermentation processes. Along heating processes, starch will gelatinize and the granule will swell and then another part of dissolved starch will influence level of viscosity. A rigid structure is difficult to swell due to a solid matrix causing limited water absorption.

### Solubility

The solubility percentage of non fermented Nagara bean flour was 20.13%db whereas the soluble fermented Nagara bean flour tended to decrease together with an increase of fermentation periods by *L. plantarum*. The solubility is explained Table 1. Lactic acid bacteria produce cellulose enzymes degrading cellulose of starch cell walls and breaking cell walls, then, granular starch will leave contributing toward the in/outflow of water and amylosa in granular starch. Due to polar characters of amylosa, the solubility will enhance;

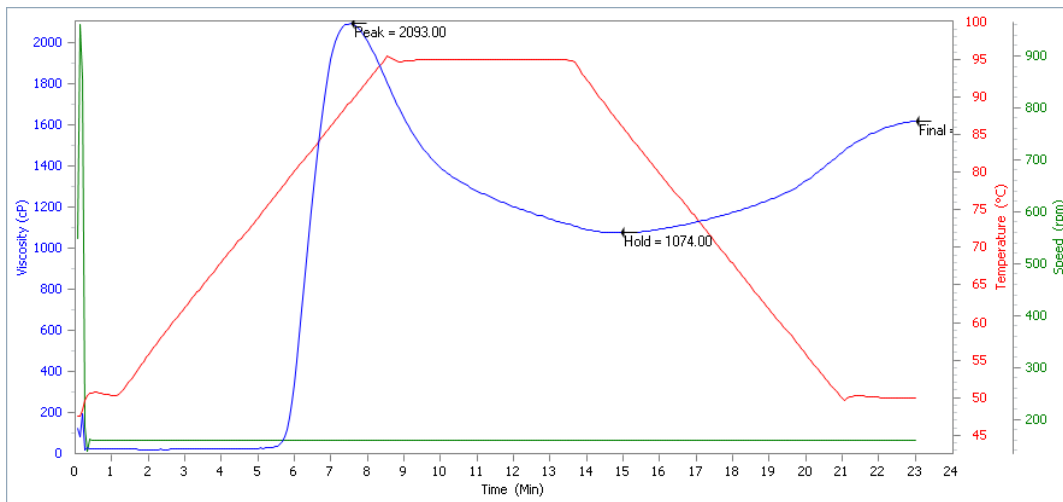


Figure 1. Gelatinization profiles of Nagara bean flour (control)

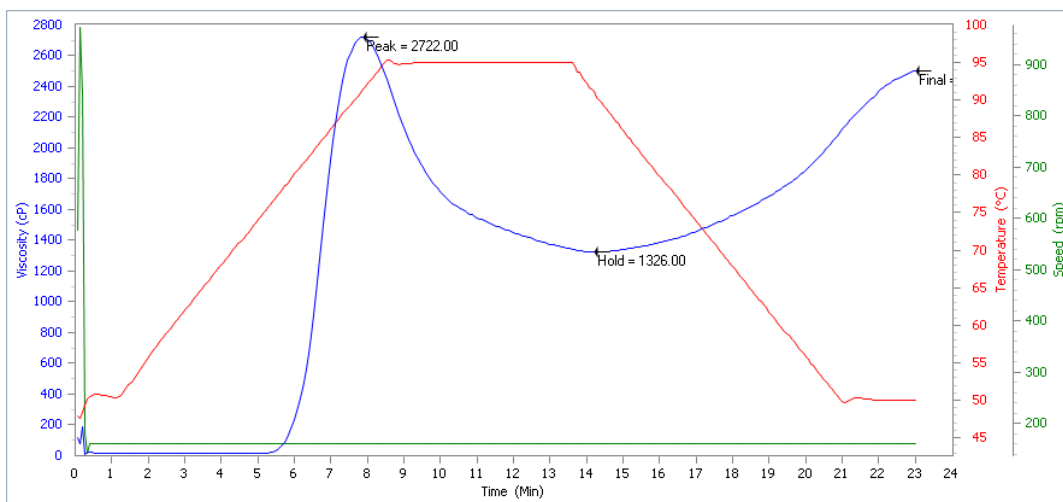


Figure 2. Gelatinization profiles of fermented Nagara bean flour using *L. plantarum* for 48 hours and grits sizes.

however, it will decline when adding fermentation periods.

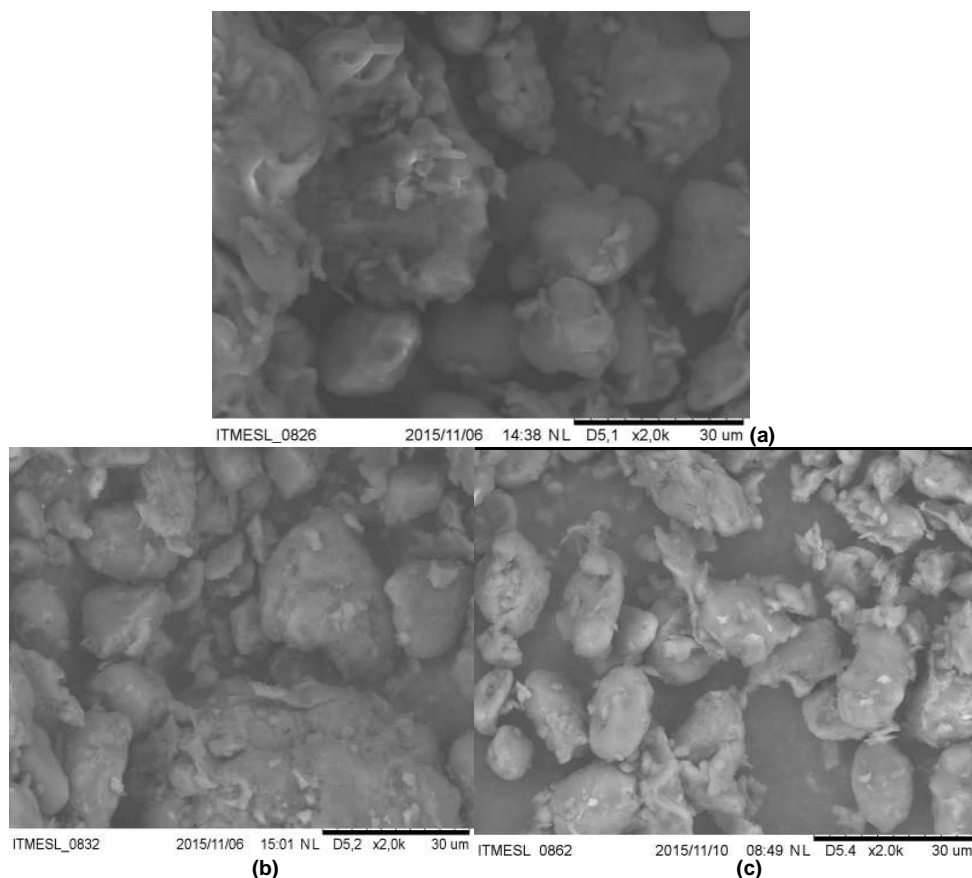
High quality of starch is characterized by an increase of starch content and produced pasta contains a low solubility, an increased volume and swelling power. The high solubility and low swelling volume show a low quality of starch because of a dilute solution and unstable pasta when it is cooked (Leach and Scoch, 1996; Afoawka and Aidoo, 2005). The strength of swelling volume and solubility of starch indicates covalent bonds between starch molecules. The ratio between amylosa-amylopectin, long chains and distributing molecule weights, conformation and degree of branched molecules play an important role in swelling volume and solubility

(Moorthy, 2001). The high amylosa and tied intramolecular contents will reduce swelling volume.

### Gelatinization profiles

A gelatinization profile of starch varies in granular characteristics and this indicates the prediction of functional properties and applicable potencies. Some changes are discovered along heating water-starch such as swelling volume, an increase of viscosity, tranlusen, solubility and losing birefringence, the changes is called gelatinization.

The high initial gelatinization temperature indicates resists granules to be enlarged. One of pasta



**Figure 3.** Scanning electron Microscopy (SEM) of granula in 2,0k x 30µm a) control; b) whole size-fermented flour by *L. plantarum*; c) grits size-fermented flour by *L. plantarum*

characteristics, pasta temperature, shows a minimum temperature to cook, energy costs and other stability of components. Peak viscosity indicates the characteristics of starch bonded water. Final viscosity is defined starch qualities, stability of products, and ability of formed pasta or gel after cooling periods. Gelatinization profiles of Nagara bean flour is described in Table 2 and Figure 1 and 2.

Gelatinization profiles of fermented Nagara bean flour were higher than the control and the fast result was showed for 48 hours of fermentation and grits size. These characters were caused by enzymes from LAB and a high lactic acid triggering the changes of starch.

Peak viscosity is the ability of starch to swell before physical changes happens (Sanni *et al.* 2004). The high peak viscosity of Nagara bean flour was noted for 48 hours fermentation and grits size showing 2713 cP (226.08 RVU). Based on Osungbaro (1990), the high value of peak viscosity indicated the high starch contents and related to water bonded starch capacities (Adobewale *et al.* 2005), then starch content of spontaneous fermented Nagara bean flour was around 50.13 – 71.97% db (Susi *et al.* 2015). Having high peak

viscosity of Nagara bean flour is suitable for some products needing high gel powers and elastic. Holding viscosity is the minimum value of stable temperatures and used to measure pasta from the breakdown along cooling periods, with the top trough value 1326 cP (110.5 RVU) from fermented Nagara bean flour for 48 hours fermentation and grits size.

Fernande and Berry (1989) stated the stability of starch from breakdown viscosity. The value of breakdown viscosity was 708 – 1396 cP (59 – 116.33 RVU). Final viscosity indicated viscosity changes after 50°C with the value of 1668 - 2906 cP (139 – 242.17 RVU). Final viscosity is generally used to determine quality of starch products forming viscous pasta or gel after cooking and cooling periods and in line with a resist pasta from friction along stirring.

Pasta temperature did not significantly differ from the control with the value of 78°C. Eniola dan Delarosa (1981) stated that pasta temperatures indicated time of a gelatinization process which meant an increase of viscosity for the first time and an early changing index due to swelling of starch. Pasta temperatures were related to water binding capacity that an increase of

temperature had results of high water binding capacities, of high gelatinization processes and of low swelling volume starch due to an increase in association levels of granular starch (Eniola and Delarosa, 1981; Numfor *et al.* 1996).

Legume starch had implied high viscosity rather than grain flour (Lineback and Ke 1975) that meant legume starch more resistant in terms of swelling and friction. The structure and size of granular starch, a degree of crystallization, protein and fat, a size of molecule and a number of branches in starch fraction influenced viscosity (Schoch and Maywald 1968). In addition, gelatinization processes was affected by starch types, granular sizes, concentration, ratio between amyloza and amylopectin, an endogen component such lipid and other additional materials. Susi *et al.* (2015) discovered level of amyloza in non fermented Nagara bean flour was lower than in fermented one using *L. plantarum* which was around 20.5% and 20.66% - 26.96% respectively. The finding indicated fermented Nagara bean flour having a high temperature of gelatinization processes.

An average of protein content in fermented Nagara bean flour was 20.5% that interrupting the process of enlarged granular starch and reducing viscosity (Hamaker and Griffin 1993; Yang and Chang 1999). Lipid and protein contents have formed inclusion complexes of amylose in native starch and flour causing decrease in rate of swelling.

Interaction between protein and starch was caused by different charges of molecule bonds and then forming inclusion complexes during gelatinization processes disturbed swelling process. Amyloza functions as diluents and inhibitors swelling processes particularly when available lipid triggered non dissolved complexes to amyloza. Starch molecule has hydrogen bonds in crystal bundle forms influencing granular bonds during swelling processes and solubility (Leach *et al.* 1959). When granular starch was heated, its structure would be hydrated and increased for high swelling, moderate swelling, restricted swelling or highly restricted swelling. Crosslink happened in granular starch would reduce swelling and stabilize the friction (Galves and Resurreccion 1993).

### Scanning electron microscopy (SEM)

Physical and chemical changes of fermented flour can be seen from its structure. In general, fermented flour will lose the soft structure of flour. Adding microbial starter will form irregular structures. Scanning electron microscopy of granular starch in grits size shows smaller size and irregular structure compared to granular starch in whole size. Granular starch in a spontaneous fermentation process and in lactic acid bacteria could be seen in Figure 3.

Figure 3 describes a small damaged level of granules; however, fermented flour both whole and grits sizes shows intensive damage. Moreover, degree of damaged granules randomly happens not in all surface of its structure.

### CONCLUSION

Fermented Nagara bean flour by *L. plantarum* showed better water absorption, swelling volume and gelatinization characteristics along 48 hours fermentation and in grits size compared to the control. Grits sizes provided faster changing rather than whole sizes and time of fermentation by *L. plantarum*.

### ACKNOWLEDGEMENT

The authors would thank to DP2M DIKTI for the finance through Hibah Pekerti in the year of 2015.

### REFERENCES

- Adebowale AA, LO Sanni, SO Awonrin. (2005). Effect of texture modifiers on physicochemical and sensory properties of dried fufu. *Food Sci Technol. Int.* 11:373-382
- Afoakwa EO, Aidoo R (2005). Changes in souring development, nutritional and functional properties during fermentation of cowpea-fortified nixtamalized maize. *Inter J Food Sci Nutr* 61: 256-271.
- Atuobi C, Sakyi-Dawson E, Sefa-Dedeh S, Afoakwa EO, Budu AS. (2011). Microstructural and Physico-Functional Characterization of Starches from Selected Cowpea (*Vigna unguiculata L. Walp.*) Varieties Developed for Pest and Disease Resistance. *J Nutr Food Sci* 1:104.
- Chinsamran K, Piyachomkwan K, Santisopasri V, Sriroth K. (2005). Effect of Lactic Acid Fermentation on Physico-chemical Properties of Starch Derived from Cassava, Sweet Potato and Rice. Thailand
- Eniola L, LC Delarosa. (1981). Physicochemical characterizes of yam starches. *J. Food Biochem.* 5:115-130
- Etudaiye HA, Nwabueze, TU, Sanni, LO. (2009). Quality of fufu Processed from Cassava Mosaic Disease (CMD) Resistant Varieties. *African Journal Of Food Science* 3 (3):061-067
- Femande De Tonella ML, JW Berry. (1989). Rheological properties of flour and sensory characteristics of bread made from germinated chick peas. *Internal. J. Food Sci. Technology*
- Galvez FCF, Resurreccion AVA (1993). The effects of decortication and method of extraction on the physical and chemical properties of starch from mung bean (*Vignaradiate (L.) wilczec*). *Journal of Food Processing and Preservation*, 17: 93-107.
- Hamaker BR, Griffin VK. (1993) Effect of disulfide bond-containing protein on rice starch gelatinization and pasting. *Cereal Chemistry*, 70:377-380.
- Leach HW, McCowen LD, Schoch TJ (1959). Structure of the Starch Granule. 1. Swelling and solubility patterns of various starches. *Cereal Chem.* 36:534-544
- Leach HW, Schoch TJ. (1996). Structure of the starch granule II. Action of the various Amylases on Granular Starches. *Cereal Chem* 38: 34-46.
- Lineback DR, Ke CH (1975). Starches and low-molecular weight carbohydrates from chick pea and horse bean flours. *Cereal Chemistry*, 52:334-347.
- Moorthy SN. (2001). Tuber crop starches in Central Tuber Crops Research Institute. 2nd Edition. St Joseph's Press. India. 18: 42.

- Niba L. (2003). The relevance of biotechnology in the development of functional foods for improved nutritional and health quality in developing countries African Journal of Biotechnology 2 (12): 632-635
- Numfor FA, WM walter, SJ Schawrtz. (1996). Effect of emulsifiers on the physical properties of native and fermented cassava starches. J. Agric. Food Chem. 44:2595-2599
- Olapade AA, OB Oluwole, OC Aworh(2012). Physico-chemical properties and consumer acceptance of instant cowpea (*Vigna unguiculata*) powder for complementary food. African J. Food Sci. Technol., 3: 102-106
- Osungbaro TO (1990). Effect of differences in variety and dry milling of maize on texture characteristics of ogi. J. Food. Agric. 54:1-11
- Sanni LO, Kasoko AA, Adebawale RJ Adeoye.( 2004). The Influence of palm oil anf chemical modification on the pasting and sensory properties of fufu flour. Int. J.Food Properties, 7:229-237
- Schoch TJ, Maywald EC(1968). Preparation and properties of various legume starches. *Cereal Chemistry*, 45:564-573.
- Susi S ( 2012). Komposisi Kimia dan Asam Amino Tempe Kacang nagara. *Agroscentiae* 19 (1) p 28-36
- Susi S ( 2015). Modifikasi Tepung Kacang Nagara Melalui Fermentasi Bakteri Asam Laktat untuk Produksi Beras Analog Dalam Rangka Ketahanan Pangan. Laporan Penelitian. Banjarbaru
- Tester, RF,WR Morrison (1990). Swelling and Gelatinization of Cereal Starches. II Waxy Rice Starches. *Cereal Chem.* 67(6):558-563
- Thangadurai D. (2005). Chemical Composition And Nutritional Potential Of *Vigna Unguiculatasp.Cylindrica* (Fabaceae). *J. Bichem.* 29 : p 88-98
- Yang CH, Chang WH. (1999). Effects of protein and lipid binding to starch on the physicochemical and pasting properties of rice flour. *J. Food Sci. and Agric. Chem.*, 1: 277- 285.